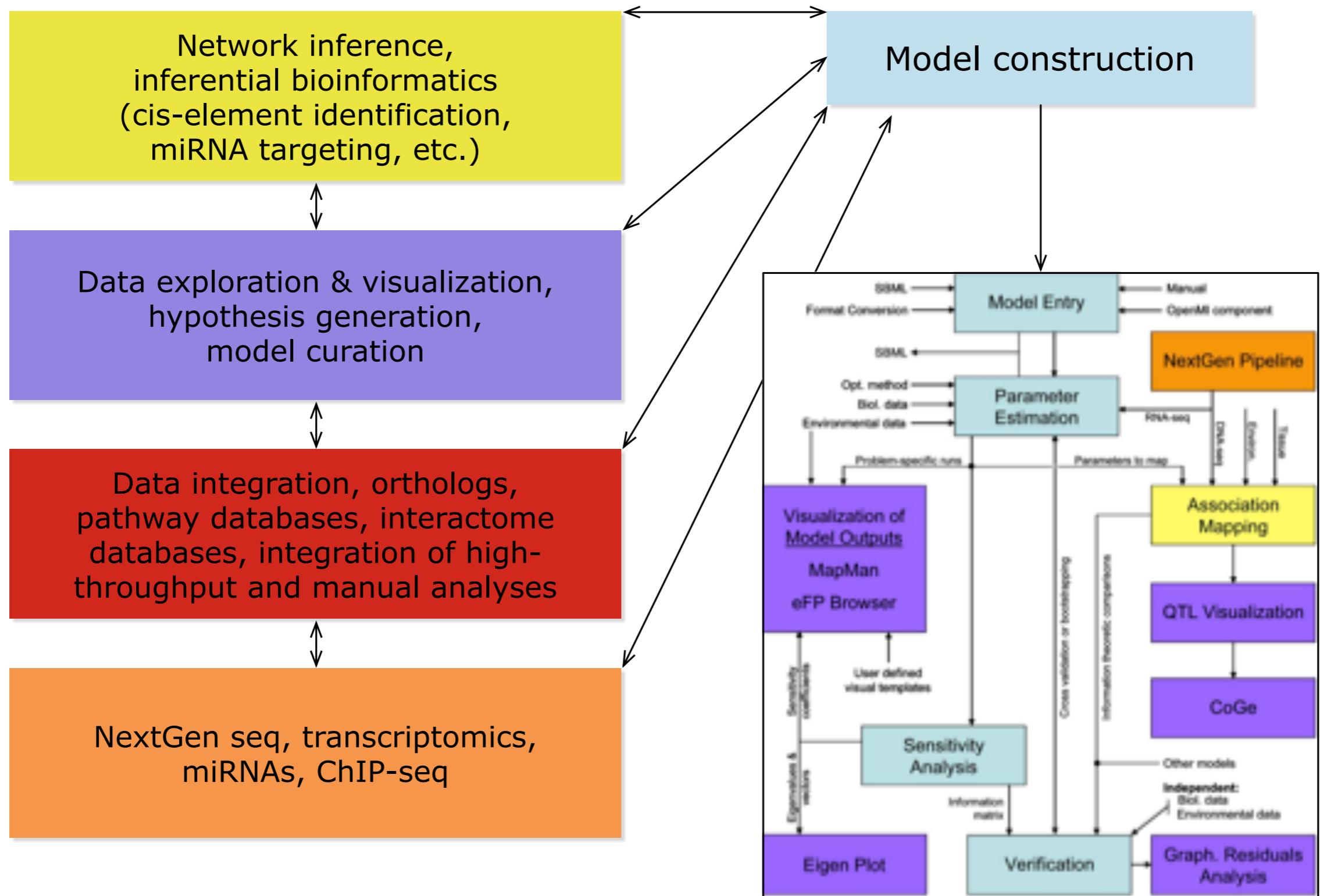


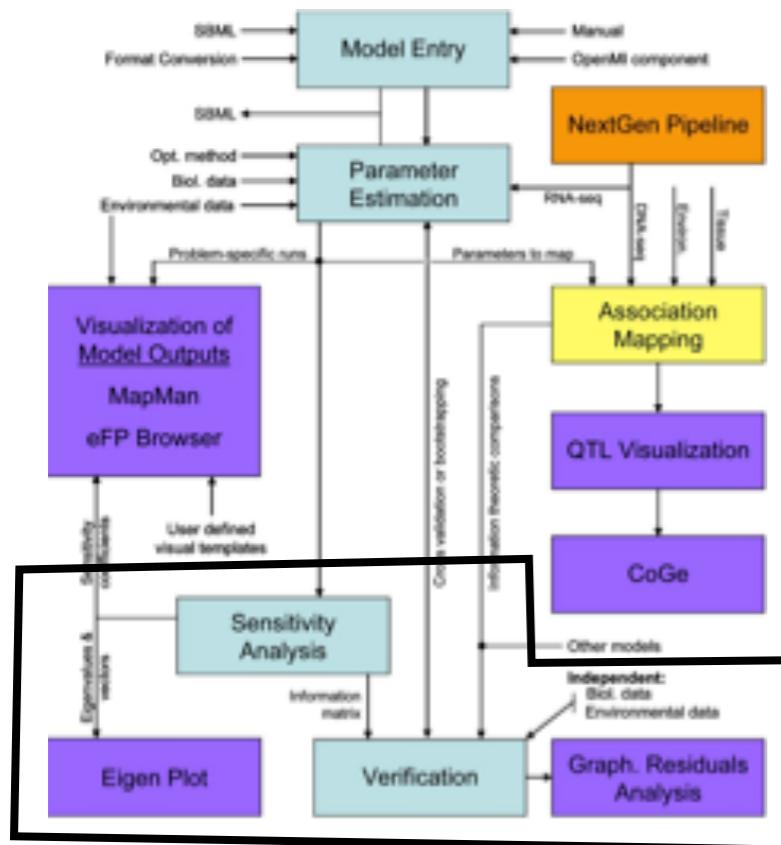
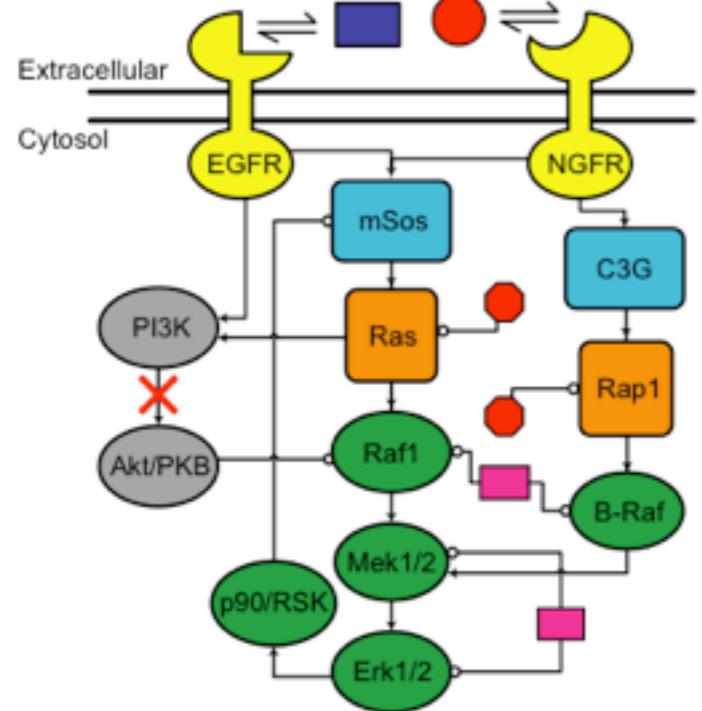
# Workflows & interactions



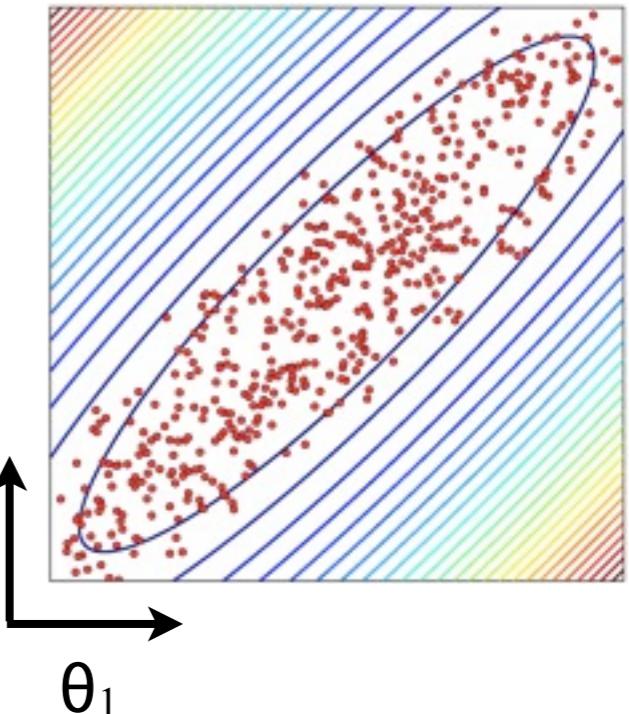
# Computational pipelines

- NextGen Pipeline(s)
  - RNA-seq, DNA-seq
- cis-element prediction
  - X. Zhu: implementing various algorithms combining sequence and expression information (de novo motif identification, over-representation of candidate elements, module networks)
  - T. Mockler: ELEMENT website implementing over-representation analysis of candidate elements (PLACE db)
- miRNA identification & target prediction
  - X. Zhu: implementing tools for (i) identifying candidate miRNAs from RNA-seq data, and (ii) identifying candidate target mRNAs
- Proposal: devote iPlant resources to support pipeline refinement, coupling, and hosting as a community resource
  - where appropriate (e.g., cis-element prediction), develop common interface and tools to assess and compare results from different algorithms

# Parameter sensitivities



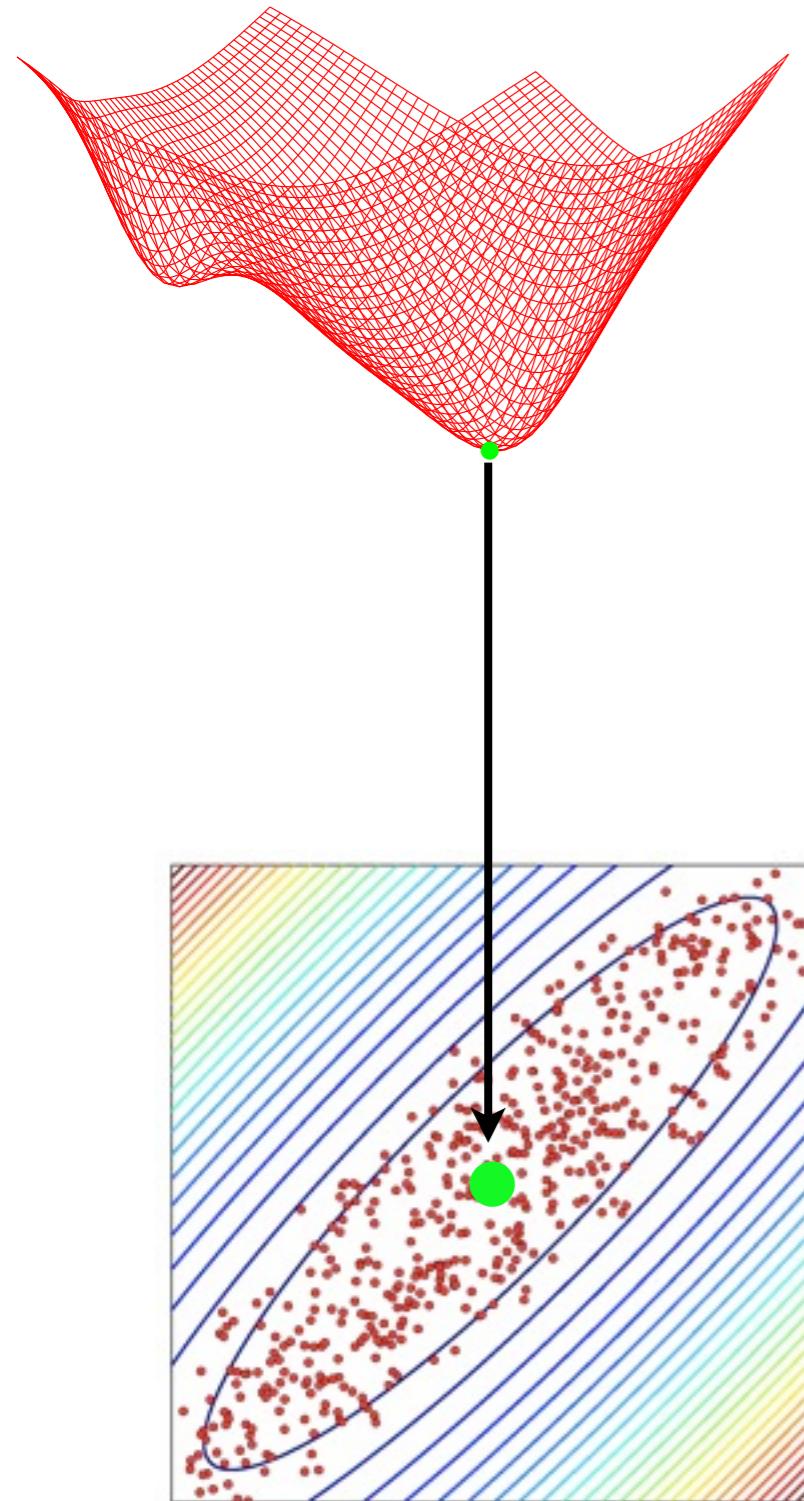
$$\begin{aligned}
 \frac{d[BGF]}{dt} &= -k_{rbEGF}[EGF][freeEGFR] + k_{ruEGF}[boundEGFR] \\
 \frac{d[NGF]}{dt} &= -k_{rbNGF}[NGF][freeNGFR] + k_{ruNGF}[boundNGFR] \\
 \frac{d[freeEGFR]}{dt} &= -k_{rbEGF}[EGF][freeEGFR] + k_{ruEGF}[boundEGFR] \\
 \frac{d[boundEGFR]}{dt} &= +k_{rbEGF}[EGF][freeEGFR] - k_{ruEGF}[boundEGFR] \\
 \frac{d[freeNGFR]}{dt} &= -k_{rbNGF}[NGF][freeNGFR] + k_{ruNGF}[boundNGFR] \\
 \frac{d[boundNGFR]}{dt} &= +k_{rbNGF}[NGF][freeNGFR] - k_{ruNGF}[boundNGFR] \\
 \frac{d[SosInactive]}{dt} &= -k_EGF[boundEGFR] \frac{[SosInactive]}{[SosInactive] + K_mEGF} \\
 &\quad - k_{NGF}[boundNGFR] \frac{[SosInactive]}{[SosInactive] + K_mNGF} \\
 &\quad + k_{dSos}[P90RskActive] \frac{[SosActive]}{[SosActive] + K_mdSos} \\
 \frac{d[SosActive]}{dt} &= +k_EGF[boundEGFR] \frac{[SosInactive]}{[SosInactive] + K_mEgF} \\
 &\quad + k_{NGF}[boundNGFR] \frac{[SosInactive]}{[SosInactive] + K_mNGF} \\
 &\quad - k_{dSos}[P90RskActive] \frac{[SosActive]}{[SosActive] + K_mdSos} \\
 \frac{d[P90RskInactive]}{dt} &= -k_pP90Rsk[ErkActive] \frac{[P90RskInactive]}{[P90RskInactive] + K_mpP90Rsk} \\
 \frac{d[P90RskActive]}{dt} &= +k_pP90Rsk[ErkActive] \frac{[P90RskInactive]}{[P90RskInactive] + K_mpP90Rsk} \\
 \frac{d[RasInactive]}{dt} &= -k_Sos[SosActive] \frac{[RasInactive]}{[RasInactive] + K_mSos} \\
 &\quad + k_{RasGap}[RasGapActive] \frac{[RasActive]}{[RasActive] + K_mRasGap} \\
 \frac{d[RasActive]}{dt} &= +k_Sos[SosActive] \frac{[RasInactive]}{[RasInactive] + K_mSos} \\
 &\quad - k_{RasGap}[RasGapActive] \frac{[RasActive]}{[RasActive] + K_mRasGap} \\
 \frac{d[RasGapActive]}{dt} &= 0 \\
 \frac{d[RafInactive]}{dt} &= -k_{RasToRaf1}[RasActive] \frac{[Raf1Inactive]}{[Raf1Inactive] + K_mRasToRaf1} \\
 &\quad + k_{dRaf1}[Raf1PPtase] \frac{[Raf1Active]}{[Raf1Active] + K_mdRaf1} \\
 &\quad + k_{dRaf1ByAkt1}[AktActive] \frac{[Raf1Active]}{[Raf1Active] + K_mRaf1ByAkt1} \\
 \frac{d[Raf1Active]}{dt} &= +k_{RasToRaf1}[RasActive] \frac{[Raf1Inactive]}{[Raf1Inactive] + K_mRasToRaf1} \\
 &\quad - k_{dRaf1}[Raf1PPtase] \frac{[Raf1Active]}{[Raf1Active] + K_mdRaf1} \\
 &\quad - k_{dRaf1ByAkt1}[AktActive] \frac{[Raf1Active]}{[Raf1Active] + K_mRaf1ByAkt1}
 \end{aligned}$$



$$\begin{aligned}
 \frac{d[SosActive]}{dt} &= +k_EGF[boundEGFR] \frac{[SosInactive]}{[SosInactive] + K_mEgF} \\
 &\quad + k_{NGF}[boundNGFR] \frac{[SosInactive]}{[SosInactive] + K_mNGF} \\
 &\quad - k_{dSos}[P90RskActive] \frac{[SosActive]}{[SosActive] + K_mdSos}
 \end{aligned}$$

*How does the behavior of a model change with changes in model parameters? (Implications for model validation, experimental design, evolution, control, etc.)*

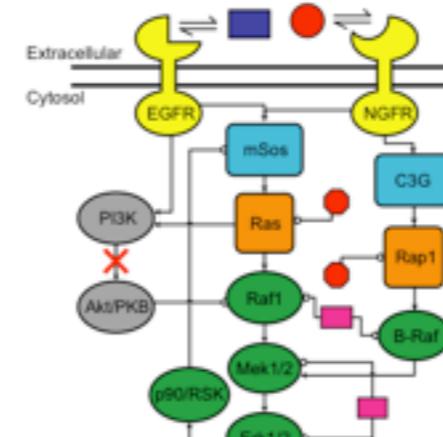
# Fits, ensembles, and uncertainties



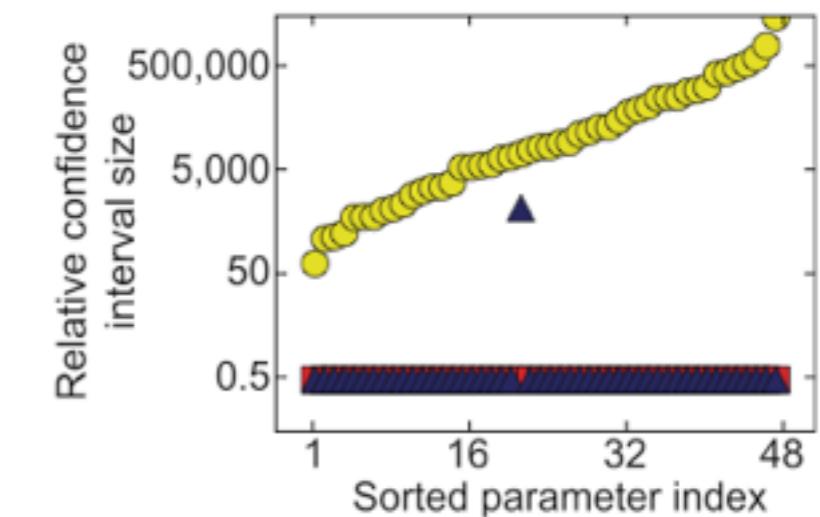
Chemical species  $y(\vec{\theta}, t)$   
Parameters  $\vec{\theta}$

Data  $y_i \pm \sigma_i$   
Cost  $C(\vec{\theta})$

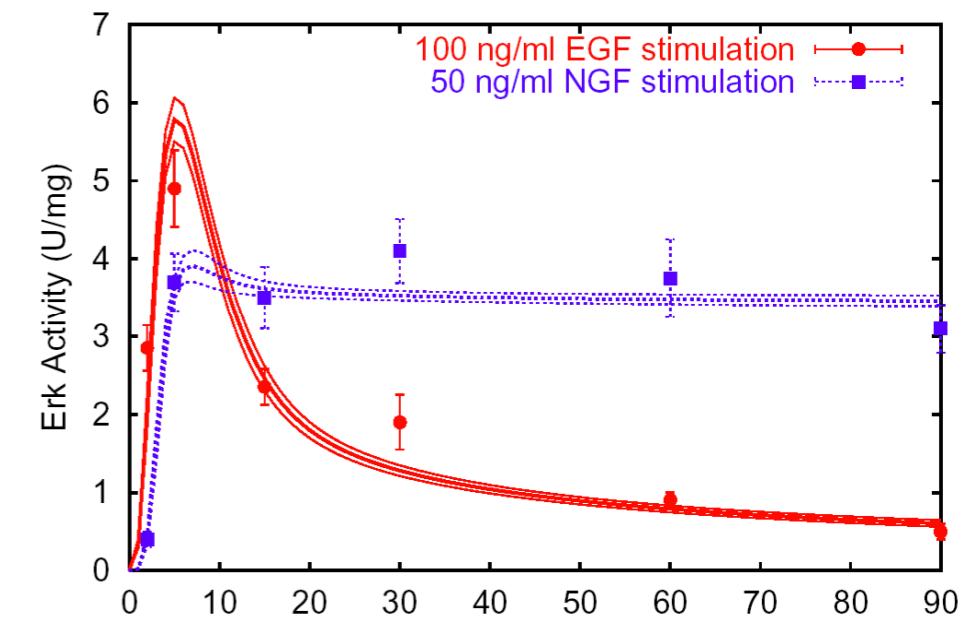
$$C(\vec{\theta}) = \frac{1}{2} \sum_{i=1}^{N_D} \frac{(y(\vec{\theta}) - y_i)^2}{\sigma_i^2}$$



Prediction uncertainties  
(from linearized analysis or  
sampling of parameter  
ensembles)

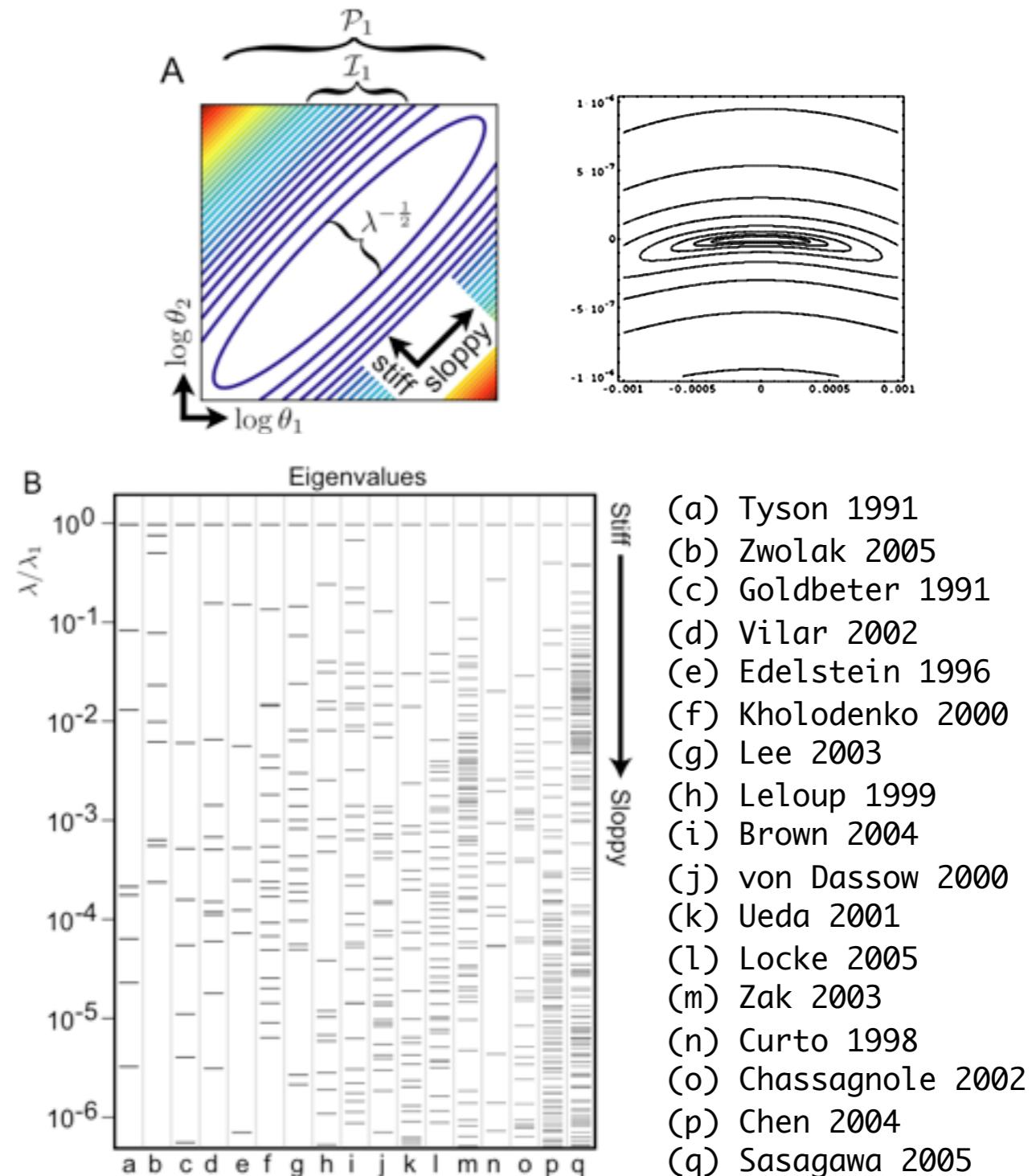


tight predictions despite  
underlying parametric  
uncertainty



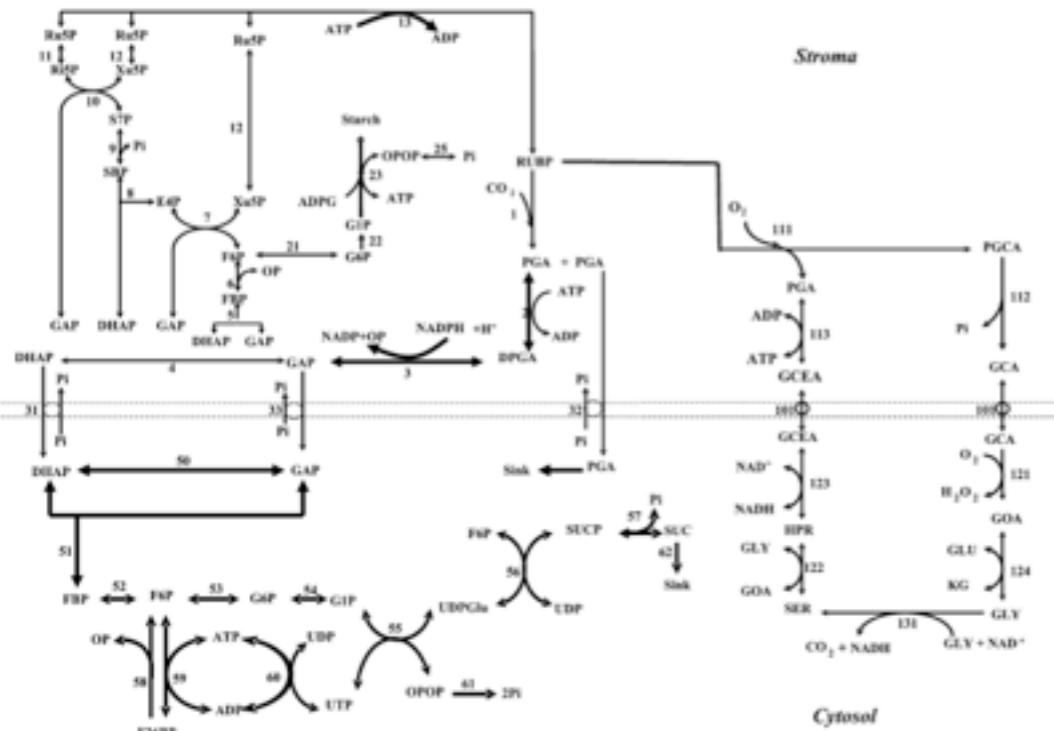
# Sloppy parameter sensitivities

- Behavior of model is typically **orders of magnitude more sensitive** to moves along some directions in parameter space than others
  - “stiff” vs. “sloppy”
- Characterized by the eigenvalues of the Hessian matrix about the best fit
  - eigenvalues span many orders of magnitude (ill-conditioned)
  - roughly constant density in  $\log \lambda$ : *a few stiff modes, and many sloppy ones*
  - similar for  $\chi^2$  analysis of “complete data” generated by model with reference parameters
  - ubiquitous across systems biology models
- Implications for experimental design
  - don’t focus on parameter identification



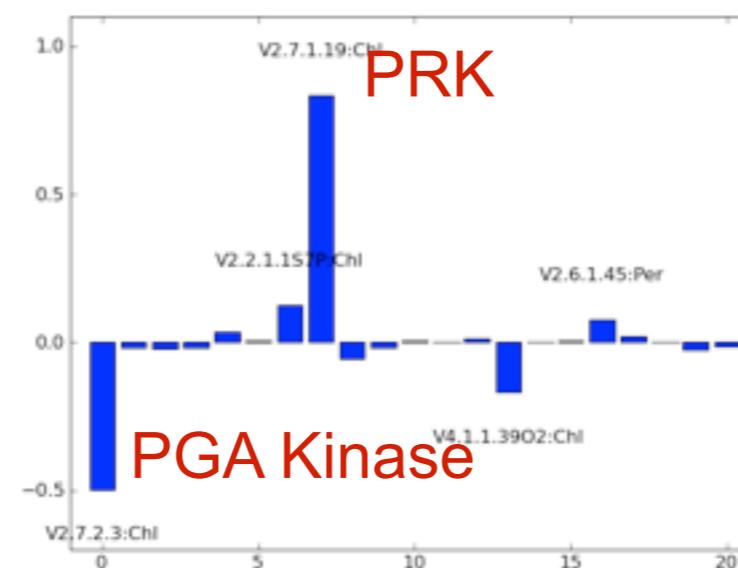
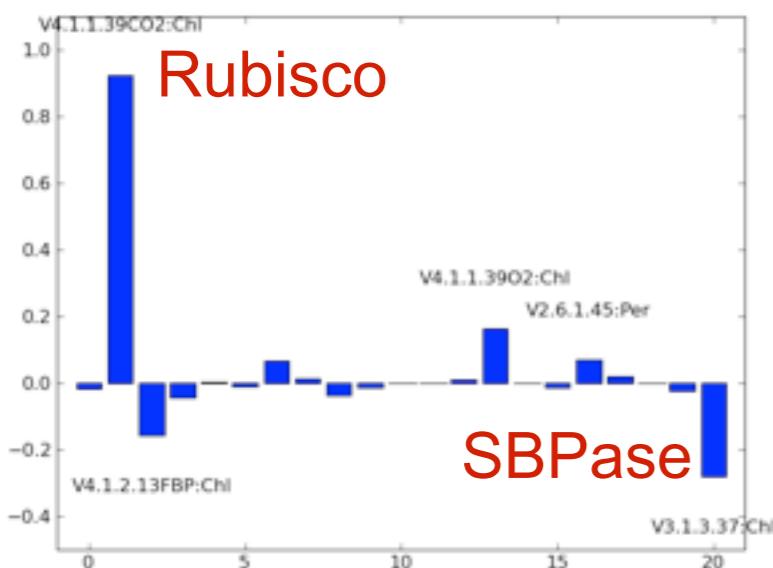
R.N. Gutenkunst *et al.*, PLoS Comp. Bio. (2007)

# Evolvability of photosynthetic metabolism



- C3 photosynthesis model (Zhu 2007): evolutionary optimization of CO<sub>2</sub> assimilation by reallocating enzyme concentrations
- examining “stiffness” and “sloppiness” of collective parameter sensitivities to probe structure of evolutionary path
- need for visualization tools to see dynamics and sensitivity analysis in context of pathway diagram
- need for tools/formats for model & simulation exchange and validation (a la BioModels.net)

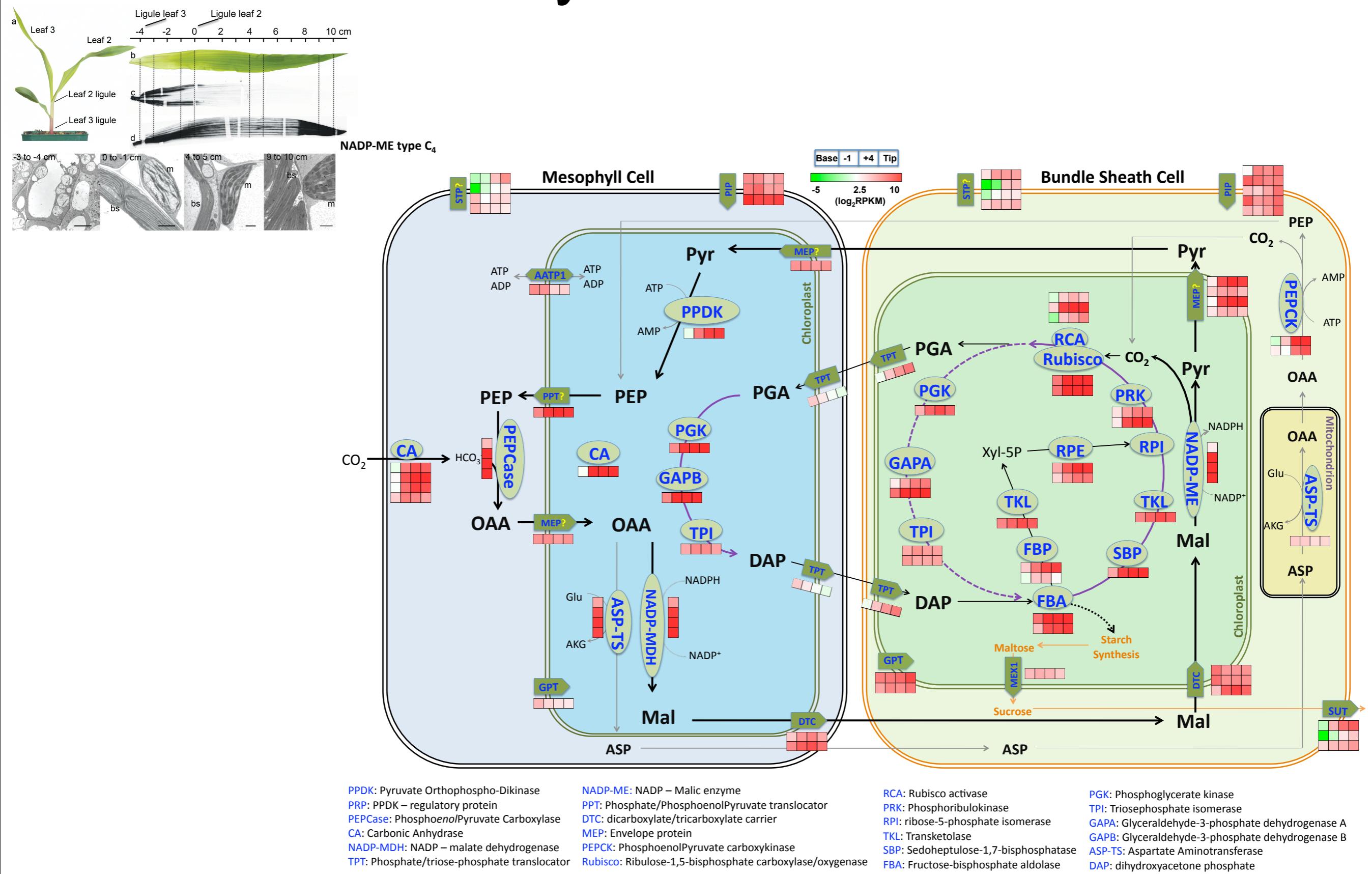
three “stiffest” parameter directions from preliminary sensitivity analysis (“Eigen Plot”)



# SloppyCell

- open-source software system for kinetic modeling, parameter estimation, uncertainty quantification and sloppy modeling
  - [sloppycell.sourceforge.net](http://sloppycell.sourceforge.net); built largely on Python & SciPy (for numerics)
- SBML read & write: allows for analysis of a broad range of published models
- symbolic engine for synthesizing model ODEs and parametric sensitivity equations from network topology and kinetic expressions
- multiple algorithms for least-squares parameter fits to data (gradient-based, gradient-free, information geometry)
- sensitivity analysis (linearized, and via Monte Carlo sampling of parameter space)
- SloppyCell in its entirety, or in pieces carved off as separate libraries, are available for further development in iPlant

# Pathways & visualization



PPDK: Pyruvate Orthophospho-Dikinase  
PRP: PPDK – regulatory protein  
PEPCase: Phosphoenol/Pyruvate Carboxylase  
CA: Carbonic Anhydrase  
NADP-MDH: NADP – malate dehydrogenase  
TPT: Phosphate/triose-phosphate translocator

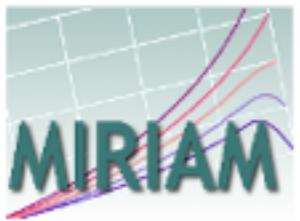
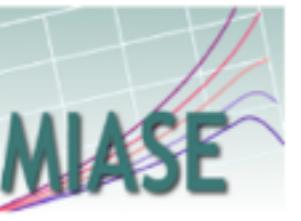
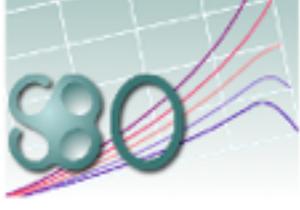
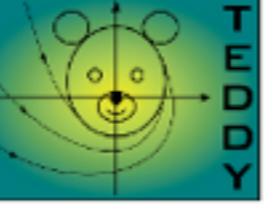
NADP-ME: NADP – Malic enzyme  
PPT: Phosphate/PhosphoenolPyruvate translocator  
DTC: dicarboxylate/tricarboxylate carrier  
MEP: Envelope protein  
PEPCK: PhosphoenolPyruvate carboxykinase  
Rubisco: Ribulose-1,5-bisphosphate carboxylase/oxygenase

RCA: Rubisco activase  
PRK: Phosphoribulokinase  
RPI: ribose-5-phosphate isomerase  
TKL: Transketolase  
SBP: Sedoheptulose-1,7-bisphosphatase  
FBA: Fructose-bisphosphate aldolase

PGK: Phosphoglycerate kinase  
TPI: Triosephosphate isomerase  
GAPA: Glyceraldehyde-3-phosphate dehydrogenase A  
GAPB: Glyceraldehyde-3-phosphate dehydrogenase B  
ASP-TS: Aspartate Aminotransferase  
DAP: dihydroxyacetone phosphate

# Standards & formats for modeling

from BioModels.net

<i>Standard specification of quantitative models</i>	<i>Model description</i>	<i>Simulation description</i>	<i>Simulation results description</i>
<i>Minimal requirements</i>	 <b>MIRIAM</b>	 <b>MIASE</b>	
<i>Data format</i>	 <b>SBML</b>  <b>SBGN</b>	<b>SED-ML</b>	<b>SBRML</b>
<i>Ontologies</i>	 <b>SBO</b>	 <b>KiSAO</b>	 <b>TEDDY</b>

- MIRIAM: Minimum Information Required in the Annotation of Models
- MIASE: Minimum Information About a Simulation Experiment
- SBML: Systems Biology Markup Language
- SBGN: Systems Biology Graphical Notation
- SED-ML: Simulation Experiment Description Markup Language
- SBRML: Systems Biology Results Markup Language
- SBO: Systems Biology Ontology
- KiSAO: Kinetic Simulation Algorithm Ontology
- TEDDY: Terminology for the Description of Dynamics

Proposal: Devote iPlant resources to determine if:

- (a) these standards are useful in supporting model validation, comparison and coupling,
- (b) there are existing tools, or a need for further tool development, to support effective use of these standards and formats,
- (c) these standards are applicable beyond chemical kinetics, e.g., to ecophysiology (or epidemiology)